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Revision: .01	Replaces: IVR/DOR1, October, 1998	Effective: 7-26-02

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A. INTRODUCTION

1. Theory

Moxidectin, ivermectin and doramectin are potent anthelmintics used in food animals to control parasitic infections. Moxidectin, ivermectin and doramectin extracted from tissue with acetonitrile; extraneous substances are removed using alumina chromatographic cleanup. All analytes are determined by HPLC after formation of a fluorescent derivative product with trifluoroacetic anhydride/1-methylimidazole. Action levels vary with species/tissue. The method is rapid, has high extraction efficiency and produces derivatized extracts suitable for direct analysis by HPLC.

2. Applicability

Tissues/species of interest are liver and muscle in bovine, ovine, porcine, caprine and equine species. This method meets sensitivity requirements and has been validated for all species/tissue combinations.

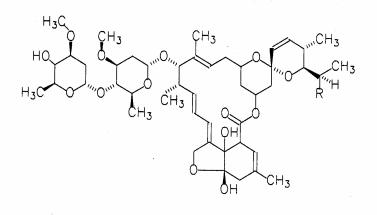
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2. Structures

Doramectin Derivative
$$\begin{array}{c} H_3C \\ H_3C \\$$

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2. Structures (cont.)



Abamectin Abamectin

(¢.

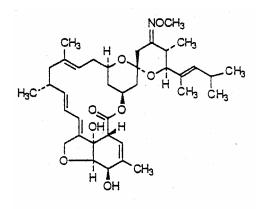
 $R = C_2H_5$ for H_2B_{1A} $R = CH_3$ for H_2B_{1B}

Abamectin Derivative

 $R = C_2H_5$ for H_2B_{1A} $R = CH_3$ for H_2B_{1B}

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2. Structures (cont.)



Moxidectin

B. EQUIPMENT

1. Apparatus

- a. N-EVAP model 112, Organomation Assoc. Inc., Berlin MA 01503.
- b. Centrifuge Sorvall model T-6000B, DuPont Co., Newton, CT 06470.
- c. Mechanical shaker Eberbach model 610 equipped with shaker box model 6040. Thomas Scientific, Swedesboro, NJ, 08065-0099.
- d. Vortex mixer Fisher Scientific, Fisher Scientific, Norcross, GA, 30091.
- e. Extraction columns Fisher Scientific Prep Sep-R (empty) Cat. No. P449R, Fisher Scientific, Norcross, GA, 30091.
- f. 50 mL screw cap centrifuge tubes Fisher Scientific #05-558-12B, Fisher

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Scientific, Norcross, GA, 30091.

- g. 50 mL polypropylene centrifuge tubes Evergreen Scientific 222-3937-G80, Evergreen Scientific International Inc., Los Angeles CA, 90058-0248.
- h. EDP Plus Micropipet, Rainin Instruments Inc., Emeryville, CA.
- i. Eppendorf Pipettor 4789, Brinkman Instrument Inc., Westbury, New York.
- j. Eppendorf Combitips 5.0 mL and 12.5 mL, Brinkman Instrument Inc., Westbury, New York.
- k. SPE Cartridges Place a small silanized glass wool plug into the neck of a 5.75" disposable transfer pipet. Add 0.1 ±0.01 g C18 bulk packing material into the disposable pipet. Tap gently to settle.

NOTE: An equivalent apparatus may be substituted for any of the apparatus above.

2. Instrumentation

- a. Liquid Chromatographic System
 - i. Waters Model 510 HPLC Pump, Waters Associates, Milford, MA.
 - ii. Waters model 717 WISP injection system, Waters Associates, Milford, MA 01757.
 - iii. Waters column temperature control module and heater. Waters Associates, Milford, MA 01757.
 - iv. Waters 474 Fluorescence detector, Waters Associates, Milford, MA 01757.
 - v. Waters model 746 Data Module, Waters Associates, Milford, MA 01757.
 - vi. Zorbax ODS 4.6 mm x 15 cm C18 analytical column, DuPont Co., Wilmington, DE 19898.
 - vii. Brownlee Labs Spheri-5 RP-18, 30 mm x 4.6 mm 5 micron guard column, Applied Biosystems, Inc. Foster City, CA 94404.
 - viii. Alltech Solvent Recycler 2000.

NOTE: An equivalent instrument may be substituted for any of the above.

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C. REAGENTS AND SOLUTIONS

- 1. Reagents and Solutions
 - a. Acetonitrile LC Grade
 - b. Alumina-Neutral type WN-3, Activity grade 1, Sigma Chemical Co., St. Louis, MO 63178. Dry at 135 ± 5 °C for at least 24 hours prior to use.

Prepare deactivated alumina for column chromatography. Alumina should be 12% deactivated. For example: Add 24 g distilled/deionized water to 176 g alumina. Mix by shaking until there are no visible lumps. Store deactivated alumina at room temperature in a tightly closed container. Use within one week after opening.

Prepare alumina columns by weighing 2.0 ±0.2 grams of deactivated alumina into an empty Prep-Sep column.

- c. 1-methylimidazole redistilled (99+ %), Aldrich Chemical Co. Cat. No. 33,609-2, Aldrich Chemical Co., Inc., Milwaukee, WI 53233.
- d. Trifluoroacetic anhydride (TFA) (99+ %) Aldrich Chemical Co. Cat. No.10,623-2, Aldrich Chemical Co., Inc., Milwaukee, WI 53233.
- e. Methanol LC Grade.
- f. Derivatizing reagents
 - i) 1 -methylimidazole 1: 1 v/v 1-methylimidazole/acetonitrile.
 - ii) TFAA 1: 1 v/v trifluoroacetic anhydride/acetonitrile.
- g. Liquid chromatograph mobile phase: 97% methanol / 3% mL water v/v.
- h. Sylon CT, Supelco # 3-3065, Supelco, Inc., Bellefonte, PA, 16823-0048.
- Waters Bondapak Cl8 bulk packing material, 125 Å, 37 55 μm, Cat. No. WATO30632.
- j. Glass Wool Silane treated.

NOTE: An equivalent reagent or solution may be substituted for any of the above.

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D. STANDARDS

- 1. Source
 - a. Ivermectin standard catalog no. L-640,471-076P004
 Merck, Sharpe and Dhome
 Rahway, NJ 07065
 - Abamectin standard catalog no. L-676-863-038A003
 Merck Sharpe and Dhome
 Rahway, NJ 07065
 - c. Doramectin standard
 Pfizer
 Lee-Summit, MO 64081-2998
 - d. Moxidectin standard-catalog no. 301423 American Cyanamid Company Princeton, NJ 08543

2. Preparation of Standards

- a. Follow manufacturer's instructions accompanying standards to obtain a stock solution of approximately 125 ±1 μg/ml in acetonitrile.
- b. Make a 1:250 dilution to obtain a spiking solution of 0.5 μg/ml in acetonitrile.
- 3. Storage Conditions
 - a. Store stock solution in freezer.
 - b. Spiking solutions may be stored at room temperature.
- 4. Shelf Life Stability
 - a. Spiking solution: 90 daysb. Stock: 1 year

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E. EXTRACTION PROCEDURE

- a. Weigh 2.5 ± 0.2 gram ground tissue into a 50-ml polypropylene centrifuge tube.
- b. Add 8 ± 0.5 ml acetonitrile and vortex for 30 ± 5 seconds.
- c. Fortify each sample with 150 ± 2μL of 0.5 μg/mL (equivalent to 30 ppb) abamectin internal standard solution. Fortify moxidectin, ivermectin and doramectin recoveries with appropriate volume of 0.5 μg/mL spiking solution depending on species analyzed.
- d. Centrifuge for 3 min \pm 30 sec. at 1500 \pm 100 RPM.
- e. Pour acetonitrile eluent through deactivated alumina column and collect eluate in a 50-ml glass centrifuge tube.
- f. Repeat extraction with additional 8 ± 0.5 ml acetonitrile, centrifuge and decant through alumina column combining eluents.
- g. Evaporate acetonitrile under a gentle stream of dry nitrogen or dry air at approx. 65 ± 5 °C.
- h. Reconstitute the dried sample from step (g) using $0.5 \text{ ml} \pm 50 \mu \text{l}$ acetonitrile. Vortex to mix.
- i. (1) Muscle tissue—Add 2 ± 0.2 ml acetonitrile and proceed to m.
 - (2) Liver tissue—Prepare SPE C18 cartridge by placing a small silanized glass wool plug into the neck of a 5.75" disposable pipet. Add 0.1 ± 0.01 g of C18 bulk packing material into the disposable pipet on top of the glass wool. Tap gently to settle C18. This SPE clean-up step for liver tissue eliminates co-extractants that interfere with quantitation.
- j. Pre-wet the SPE cartridge with 1.0 ± 0.1 ml acetonitrile. Discard the wash.
- Load the 0.5-ml sample, from step h, onto the wet SPE cartridge. (The columns must not dry out at any time or the Avermectin recoveries will be low).
 Collect the eluent.
- I. Add 2 ± 0.2 mL of acetonitrile to the sample tube. Mix. Add to the SPE column. Collect the eluent in the same container as the initial 0.5 ml eluent.
 - Note: Properly dispose of the SPE column.
- m. Add 200 µl 1-methylimidazole/acetonitrile reagent a to the eluent and vortex at least 10 sec.
- n. Add 200 µl TFA/acetonitrile reagent and vortex at least 10 sec.
- o. Inject on HPLC.

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NOTE: Derivatized samples decompose on exposure to strong light. Allow

the sample to derivatize in the dark for approximately 15 minutes before

HPLC analysis.

F. ANALYTICAL PROCEDURE

1. Instrumental conditions

Note: System may be adjusted to insure optimum response.

a. Flow rate: 1.8 ml/min.

b. Column temp. 30°C.

c. Injection volume: 50 μL – As determined by detector/integrator conditions.

d. Run time: 15 min.

e. Detector settings

i. Excitation wavelength 365 ± 20 nm

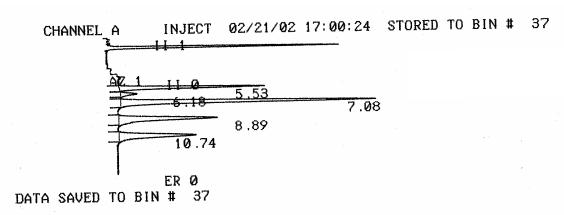
i. Emission wavelength 465 ± 20 nm

f. Detector gain: As system conditions dictate to ensure approximately ½ scale peak heights.

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2. Sample Chromatograms

a. 15 ppb Standard (Ivermectin, Abamectin, Doramectin, and Moxidectin).

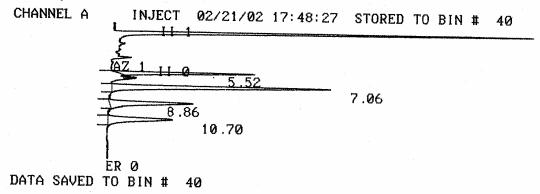


AVERMECTIN SCRE	EN	02/21/	02 17:00:24	4	CH= "A"	PS=	1.
FILE 1. ME	THOD 2.	RUN ***	INDEX	1	CALIB	BIN	37
ANALYST: DMH							
NAME	PPB	RT	PK HT BC		RF RF	T ·	
MOXIDECTIN ABAMECTIN INTE DORAMECTIN IVERMECTIN	15. RNAL STD 15. 15.	5.53 7.08 8.89 10.74	459 01 809 01 309 01 241 01		1 1	.781 .256 .517	
TOTALS	46 .		1818				

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2. Sample Chromatograms (con't)

b. 15 ppb Beef Liver Recovery (Ivermectin, Abamectin, Doramectin, and Moxidectin).



AVERMECTIN SCREEN 02/21/02 17:48:27 CH= "A" PS= FILE 1. METHOD 2. RUN *** INDEX 1 BIN 40

ANALYST: DMH

SAMPLE REC BL 1 BIN 40 NAME ARUN1734 SA IS XF 1. 1.

1.

NAME	PPB	RT	РК НТ	ВС	RF	RRT
MOXIDECTIN ABAMECTIN DORAMECTIN IVERMECTIN	15.814 INTERNAL STD 14.807 14.662	5 .52 7 .06 8 .86 10 .7	421 701 265 203	01 01	26 .643 1 . 17 .874 13 .868	0.782 1. 1.255 1.516
TOTALS	45 .283		1591		*	

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G. CALCULATIONS

Quantitation is performed by measuring peak heights. Each set is accompanied by an external standard curve at 15, 30, and 60 ppb.

Measure peak height of abamectin, ivermectin, moxidectin, and doramectin peaks in the standards and calculate the peak height ratios.

Construct a linear regression line using the ratios and standard concentrations. The correlation coefficient should be >0.995.

The equation is y = mx + b

x = Ivermectin, Moxidectin, or Doramectin /Abamectin peak height ratio

y = Ivermectin, Moxidectin, or Doramectin concentration (ppb)

m = slope

b = y-intercept

Incurred tissue ivermectin, moxidectin, or doramectin concentrations should be calculated using this regression line.

H. HAZARD ANALYSIS

- 1. Method Title DETERMINATION OF IVERMECTIN, DORAMECTIN, and MOXIDECTIN RESIDUES IN ANIMAL TISSUES.
- 2. Required Protective Equipment Safety glasses, appropriate gloves, lab coat.
- Hazards

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Reagents	Hazard	Recommended Safe Procedures
Acetonitrile, Trifluoroacetic anhydride, 1-methylimidazole	Flammable and corrrosive, may cause skin or respiratory irritation.	Avoid contact or prolonged exposure to vapors. Work in a fume hood. Keep away from flame or heat.
Ivermectin Abamectin	Weak teratogen and possible mutagen	Handle with extreme caution.
Doramectin	Severe explosion hazard if in powdered form.	Handle with extreme caution.
Moxidectin	May cause skin or respiratory irritation. The toxic effects of this material have not been fully evaluated.	Work in a well ventillated area. Store material in a secure, dry, cool well ventillated room.

4. Disposal Procedures

Dispose of solvents according to local, state, and federal guidelines.

I. QUALITY ASSURANCE PLAN

1. Performance Standard

Analyte	Analytical Range	Acceptable Recovery	Acceptable Repeatability (CV)
Ivermectin	≥ 7.5 ppb	60-120	< 20
Doramectin	≥ 7.5 ppb	60-120	< 20
Moxidectin	≥ 7.5 ppb	60-120	< 20

Regression coefficient ≥ 0.995

The Measurement Uncertainty and Method Detection Limit should be recalculated yearly

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or whenever a change that affects method accuracy, precision, or sensitivity occurs.

2. Critical Control Points and Specifications

Record Acceptable Control

a.	Sample weight	2.5 g. ± 0.2 g.
b.	Alumina deactivation level	12%
C.	1-methylimidazole volume	200 μΙ
d.	TFA volume	200 ul

3. Readiness To Perform

- a. Familiarization
 - i. Phase I:

Standards- Standard curves on each of 3 consecutive days, which will include the following:

- (a) 0 ppb
- (b) 7.5 ppb
- (c) 15 ppb
- (d) 30 ppb
- (e) 60 ppb

ii. Phase II:

Fortified samples- 3 replicates at 0, $\frac{1}{2}$ x, x, and 2x where x represents the action level for each species (bovine, ovine, and porcine) over a period of 3 different days.

NOTE: Phase I and Phase II may be performed concurrently.

iii. Phase III:

Check samples for analyst accreditation.

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- (a) 15 samples fortified between 0 and 60 ppb.
- (b) Samples submitted by the Quality Assurance Manager (QAM) or supervisor
- (c) Letter from QAM is required to commence official analysis.
- 4. Acceptability criteria.
 - i. Correlation coefficient > 0.995 for all three days.
 - ii. Mean recovery for each species in range of 60-120%.
 - iii. Repeatability for each species < 20%.
 - iv. No false positive or false negative results.
- 5. Intralaboratory Check Samples
 - a. System, minimum contents.
 - i. Frequency: One check sample per week per analyst as samples analyzed.
 - ii. Blind check samples: Samples chosen at random by supervisor or QAM.
 - iii. Records are to be maintained by the analyst and reviewed by the supervisor and QAM for:
 - (a) All replicate findings.
 - (b) CUSUM and/or SHEWHART charts.
 - (c) All recovery values.
 - (d) Running average, standard deviation, and CV for all recoveries.
 - b. Acceptability criteria.

If unacceptable values are obtained, then:

- i. Stop all official analyses by that analyst.
- ii. Take corrective action.
- 6. Sample Acceptability and Stability
 - a. Matrices: Liver, Muscle

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b. Species: Bovine, porcine, ovine, caprine and equine.

c. Sample size: 16-oz minimum.

d. Condition on receipt in Sample Preparation Section: frozen.

e. Sample Storage:

i. Time: 6 monthsii. Condition: Frozen

7. Sample Set

- a. Standards at 15, 30 and 60 ppb.
- b. Recovery at x level.

8. Sensitivity

- a. Lowest detectable level (LDL): 2 ppb.
 - i. Lowest reliable quantitation (LRQ): 7.5 ppb.
 - ii. Minimum proficiency level (MPL): 7.5 ppb.

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J. WORKSHEET

The worksheet on the next page can be removed from this book for photocopying.

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K. References

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